





Regulation of cancer cell metabolism and proteostasis by mechanical tissue cues

Category: Experimental Hematology / Oncology

J. Hill^{1,2}, M. Román-Trufero^{1,2}, K. Blighe³, E. Gentleman^{4,5}, H.W. Auner^{1,2,6}

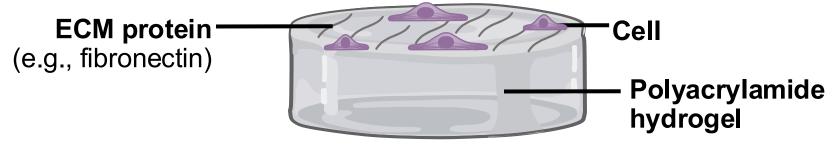
¹Service and Central Laboratory of Hematology, Lausanne University Hospital (CHUV), Lausanne, Switzerland. ²Department of Immunology and Inflammation, Imperial College London, London, United Kingdom. ³ Clinical Bioinformatics Research, London, United Kingdom. ⁴ Department of Biomedical Sciences, University of Lausanne, Lausanne, Switzerland. ⁵ Centre for Craniofacial and Regenerative Biology King's College London, Guy's Hospital, London, United Kingdom. ⁶ Faculty of Biology and Medicine, University of Lausanne, Lausanne, Switzerland.

Background and Objectives

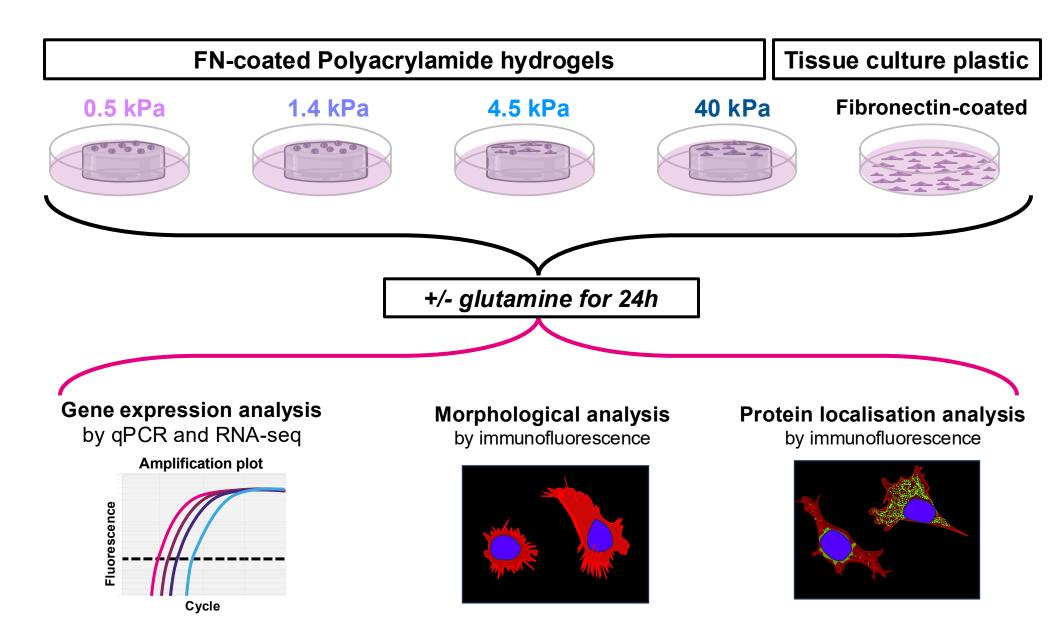
- The integrated stress response (ISR) is a signalling pathway that regulates protein synthesis in response to diverse stressors (e.g., amino acid scarcity) and is implicated in cancer progression and treatment resistance.
- Phosphorylation of the α subunit of eukaryotic translation initiation factor 2 (elF2α) attenuates global mRNA translation, while enhancing selective translation of mRNAs, including activating transcription factor 4 (ATF4). This promotes an adaptive cellular response to aid recovery and survival to re-establish homeostasis or, alternatively, induce cell death.
- Tissue **stiffness**, a mechanical property of the tumour microenvironment determined by the extracellular matrix (ECM), influences cell behaviour through mechanotransduction (e.g., via YAP/TAZ).
- Stiffer tumour tissue promotes the proliferative, invasive, and metastatic capacity of cancer cells, and promotes drug resistance by impeding drug delivery.
- Given the importance of both mechanotransduction and the ISR in cancer biology, we hypothesise that stiffness may modulate ISR activation, thereby linking mechanical cues to stress adaptation in tumor cells.

Methods

- Morphological analysis was performed to assess stiffness sensitivity of cell lines, whereby cell area (um²) and circularity was measured via immunofluorescence and quantified using Fiji software.
- Gene expression of key YAP/TAZ targets, CYR61 or CTGF, was



• ISR pathway was activated in A375 melanoma cells by glutamine starvation and assessed by qRT-PCR, RNA sequencing, and immunofluorescence to evaluate global gene expression and



Results

E = 0.2 to 1.0 kPa

Figure 1. Solid cancer cell lines exhibit stiffness-dependent changes in

Solid cancer cell lines of varied tissue origin responded to substrate stiffness

of polyacrylamide hydrogels in a similar manner; with reduced cell area and

increased cell circularity on softer substrates, as opposed to increased

Expression of key YAP/TAZ target genes, CYR61 or CTGF, increased in cells

cell area and reduced cell circularity on stiffer substrates (Fig. 1A-B).

on stiffer substrates, indicating upregulated YAP/TAZ activity (Fig. 1C).

E = 0.5 to 5.0 kPa

MiaPaCa-2

morphology and YAP/TAZ activity.

E = 368 Mpa

E = 1.0 to 3.0 kPa

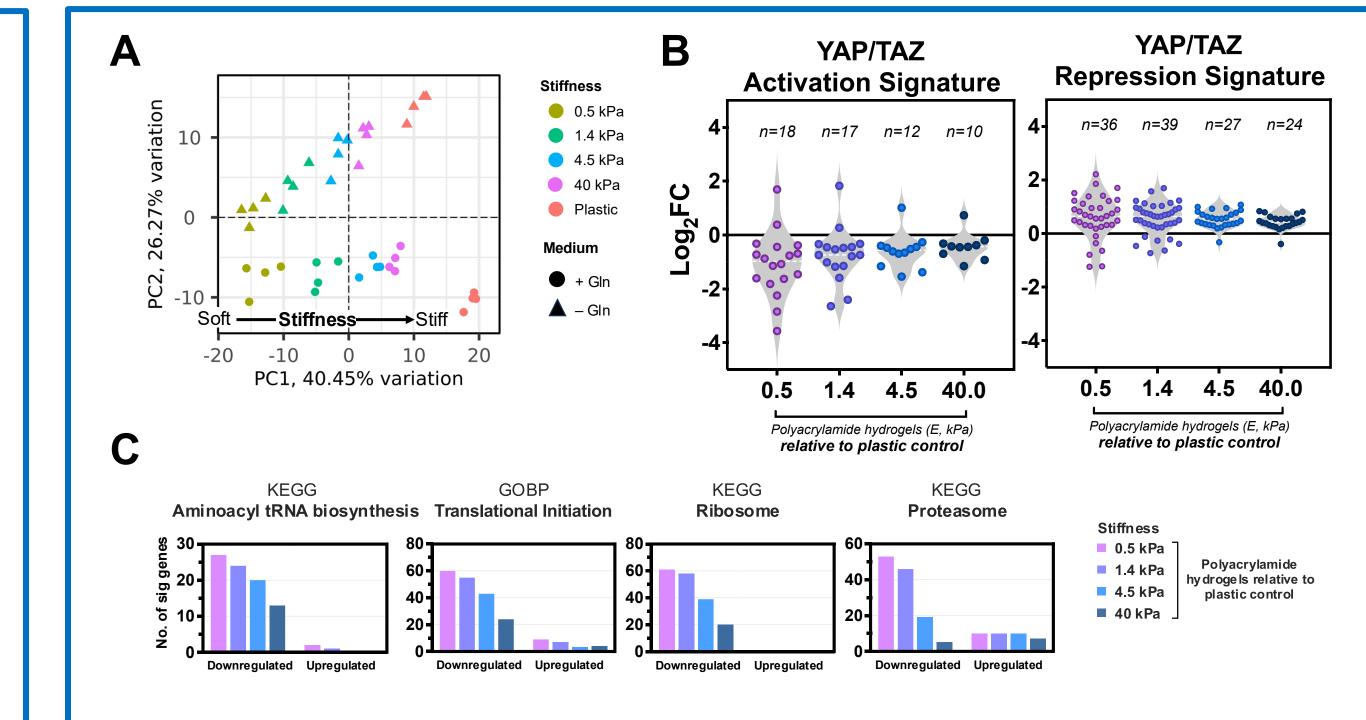


Figure 2. Softer substrates downregulate protein synthesis and degradation pathways.

- Whole transcriptome analysis of A375 cells on substrates of varied stiffness by principal component analysis revealed a progressive stiffness-dependent variation in gene expression profile both in the presence or absence of glutamine (Fig. 2A).
- Expression of a broader range of YAP/TAZ targets by RNA-seq, using 'Activation' or 'Repression' gene signatures, revealed progressively downregulated YAP/TAZ activity with reducing stiffness (Fig. 2B).
- Gene set enrichment analysis revealed that stiffness, alone, regulates a variety of protein turnover-related gene sets (Fig. 2C).

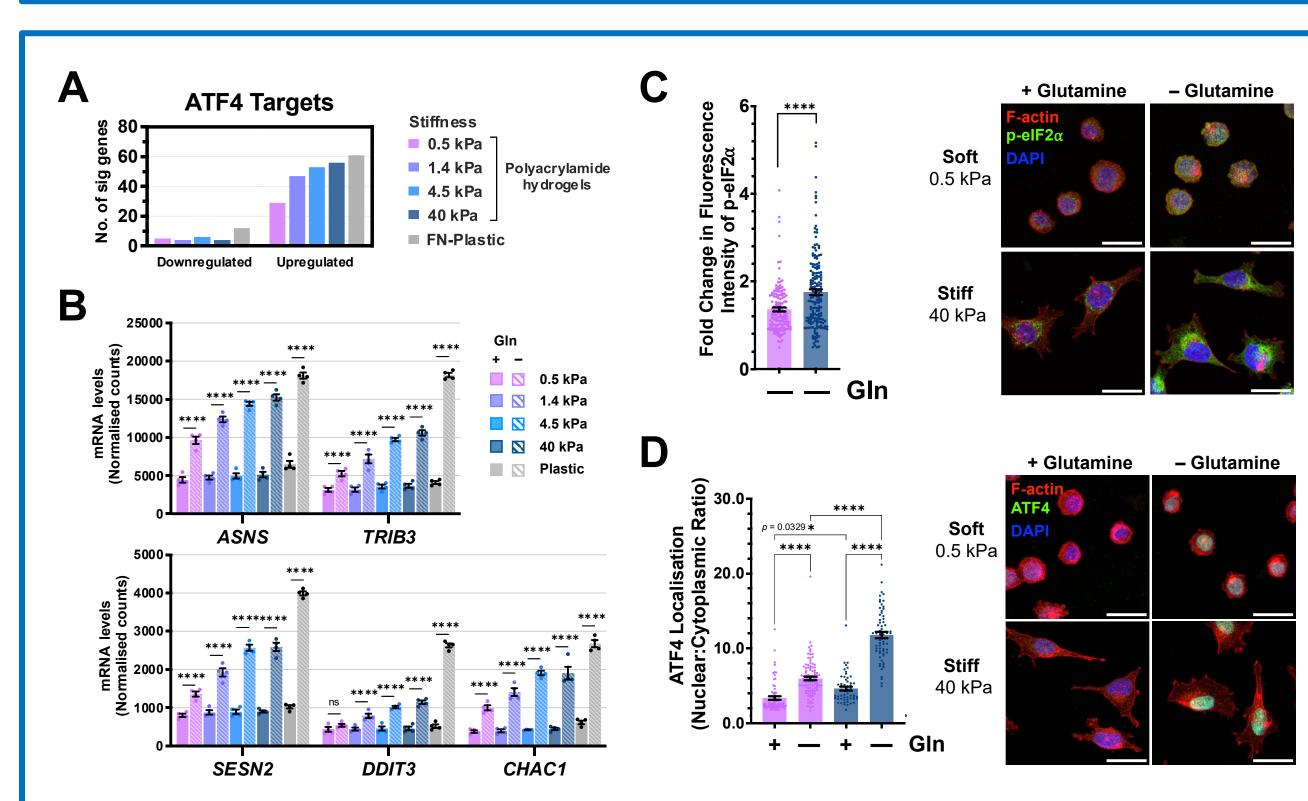


Figure 3. Extent of ISR activation induced by glutamine starvation is stiffnessdependent.

- Glutamine-starved A375 cells exhibited a stiffness-dependent difference in the expression of ATF4 targets (Fig. 3A), including expression of key ISR-activated genes (Fig. 3B).
- We examined whether the sensing or execution of the ISR was altered in A375 cells on soft (0.5 kPa) or stiff (40 kPa) polyacrylamide hydrogel substrates. This revealed increased phosphorylation of eIF2 α in cells on stiff versus soft substrates (Fig. 3C) and a higher nuclear/cytoplasmic (N/C) ratio of ATF4 in cells on stiffer substrates (Fig. 3D).

Conclusion

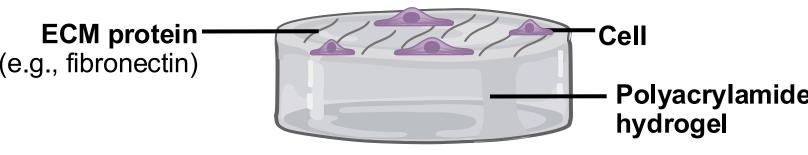
- Cancer cells of varied tissue origin respond to substrate stiffness in a similar manner, with similar morphological and transcriptional outcomes.
- Cancer cells progressively downregulate protein turnover mechanisms as substrate stiffness decreases, including those involved in translation.
- Stress sensing was increased in cells on stiffer substrates than those on softer substrates at the transcriptional level.
- Cells on softer substrates exhibited weaker ISR induction when induced by glutamine starvation.
- ISR sensing (p-eIF 2α) and execution (ATF4 nuclear localisation) were increased in cells on stiffer substrates.
- Cellular stress responses highly depend on the stiffness of the microenvironment with potential implications for cancer therapy.
- Our study highlights that caution should be implemented when interpreting cellular stress response studies in cells on tissue culture plastic due to the supraphysiological stiffness of tissue culture plastic.

Acknowledgements

This study was funded by an **UNIL-FBM Interdisciplinary Grant** to HWA and EG, the Department of Immunology and Inflammation, Imperial College London, a Cancer Research UK Advanced Clinician Scientist Fellowship to HWA, and ISREC Foundation.

Presented at the SOHC from 19 – 21 November 2025

- Solid cancer cell lines (A375, A375-MA2, MiaPaCa-2, Saos-2, and T98G) were cultured on fibronectin-coated polyacrylamide hydrogels with stiffness ranging from 0.5 to 40 kPa.
- assessed by qRT-PCR.



regulation of selected ISR and proteostasis networks.

